

1,3-DITHIANES WITH ACID FUNCTIONALITIES: POTENT INHIBITORS AND CANDIDATE AFFINITY PROBES FOR THE GABA-GATED CHLORIDE CHANNEL

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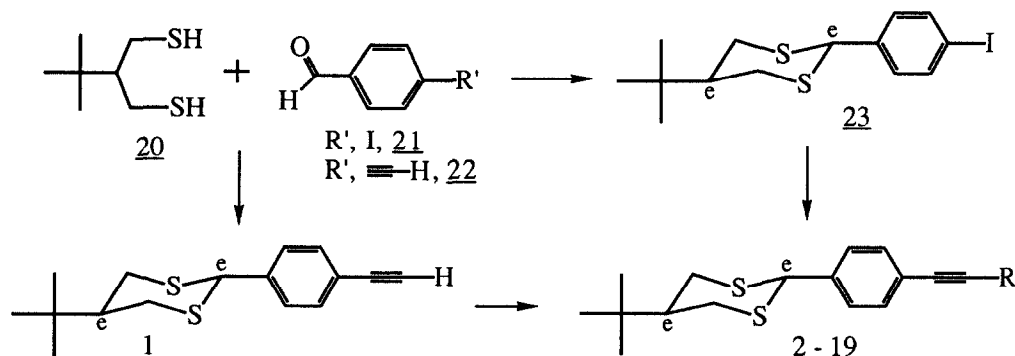
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Abstract: 2*ε*,5*ε*-1,3-Dithianes of the type $(\text{CH}_3)_3\text{C}-\text{CH}(\text{CH}_2\text{S})_2\text{CH}-\text{C}_6\text{H}_4-4-\text{C}\equiv\text{C}-\text{R}$ ($\text{R} = \text{CO}_2\text{H}$, $\text{CH}_2\text{CH}_2\text{CO}_2\text{H}$ and $\text{CH}_2\text{CH}_2\text{PO}_3\text{H}_2$) are potent blockers of the GABA-gated chloride channel with 50% inhibition at 5-10 nM. Functionalization of the acid moieties provides candidate photoaffinity ligands [$\text{R} = \text{C}(\text{O})\text{CHN}_2$ and $\text{CH}_2\text{CH}_2\text{C}(\text{O})\text{CHN}_2$], affinity columns, and hapten-protein conjugates for antibody production.

The γ -aminobutyric acid (GABA)-gated chloride channel is one of the most sensitive and important targets for pharmaceuticals, convulsants and insecticides. Characterization of this channel has been facilitated by various radioligands, photoaffinity probes, and/or affinity columns specific for the GABA, benzodiazepine, or avermectin site.¹⁻³ The classical GABA antagonist, picrotoxinin, and a variety of insecticides act primarily at the noncompetitive blocker (NCB) site.⁴ A preferred radioligand for the NCB site is 4'-ethynyl-4-[2,3-³H]propylbicycloorthobenzoate [³H]EBOB), $[\text{CH}_3\text{CH}_2\text{CH}_2-\text{C}(\text{CH}_2\text{O})_3\text{C}-\text{C}_6\text{H}_4-4-\text{C}\equiv\text{CH}]$.^{5,6} This radioligand, based on analogy with [³⁵S]tert-butylbicyclophosphorothionate [$(\text{CH}_3)_3\text{C}-\text{C}(\text{CH}_2\text{O})_3\text{P}=\text{S}$],^{7,8} measures GABA-stimulated chloride ion flux such that inhibition of radioligand binding is proportional to inhibition of chloride flux. 4-tert-Butylbicycloorthobenzoate [$(\text{CH}_3)_3\text{C}-\text{C}(\text{CH}_2\text{O})_3\text{C}-\text{C}_6\text{H}_5$] has been modified at the 1-substituent to obtain candidate photoaffinity ligands [e.g. 1-diazocyclohexadienonyl⁹ and 1-(4-azidophenyl)¹⁰] and a chemical affinity probe [1-(4-isothiocyanatophenyl)].¹¹ Although some of these orthocarboxylates are of relatively high affinity, their use is limited by acid lability and they are not practical at present.

Three recent discoveries provide an alternative approach to affinity probes for the NCB site. First, the bicycloorthocarboxylate moiety can be replaced with a dithiane with retention of much of the receptor affinity and greatly improved stability.^{12,13} Second, the dithiane is further enhanced in potency by δ -oxidation, particularly to the monosulfone.^{13,14} Third, the optimal substituents with the 1,3-dithianes are 5*ε*-tert-butyl and 2*ε*-(4-ethynylphenyl),^{12,13} the latter moiety providing a potential terminal site for functionalization. Previous additions to the ethynyl moiety in this series were not advantageous¹² but in contrast we find that this site can be extensively modified with high potency and substituents suitable for preparing affinity probes.

Synthesis of the candidate affinity probes and related compounds (**1-19**) is shown in the scheme. The intermediates were 2-*tert*-butylpropan-1,3-dithiol (**20**)¹⁵ and aldehyde **21** (Karl Industries, Aurora, Ohio) which were quantitatively converted to **23** in formic acid.¹² **21** was also used to obtain **22** by palladium-catalyzed coupling with (trimethylsilyl)acetylene followed by removal of the trimethylsilyl protective group.¹⁶



Reaction conditions and yields for the substituted-ethynylphenyl derivatives (**1-19**, see Table for R substituents) were as follows:

1: **20**, **22**, HCO₂H; 100%

2-9, **16** and **17**: **23**, excess alkyne, (Ph₃P)₂PdCl₂, CuI/Et₃N; 90-100%

10 and **12**: **a**, **14** or **16**, KOH/H₂O/MeOH/CH₂Cl₂; **b**, H⁺/H₂O; 100%

11: **15**, HCl/THF; 31%

13: **a**, **17**, Me₃SiBr; **b**, H₂O/acetone; 100%

14: **a**, **1**, MeMgCl/THF, -20 °C; **b**, ClCO₂Me/THF, -20 °C; 60%

15: **a**, **1**, *n*-BuLi /THF, -20 °C; **b**, BF₃·Et₂O, -20 °C; **c**, N₂CHCO₂Et, -20 °C; **d**, H₂O; 43%

18 and **19**: **a**, **10** or **12**, excess (COCl)₂/CH₂Cl₂/benzene, 0 °C; **b**, CH₂N₂/Et₂O, 0 °C; 5% for **18**, 36% for **19**

1 was obtained by coupling **20** and **22** according to Elliott *et al.*¹² Compounds **2-9** and **16** were prepared by reacting **23** with an excess amount of the corresponding propargyl reagent (commercially available) in the presence of bis-triphenylphosphine palladium dichloride and a catalytic amount of copper(I) iodide in anhydrous triethylamine.¹⁶ **12** was obtained from basic hydrolysis of its methyl ester (**16**). **10** was also prepared as the methyl ester (**14**) by a procedure similar to that of Earl and Townsend¹⁷ followed by hydrolysis. **15**, prepared by the general method of Hooz and Layton,¹⁸ was subjected to acid hydrolysis to give **11**. **13** was obtained via 5*e*-*tert*-butyl-2*e*-[4-(diethyl butyn-3-ylphosphono)phenyl]-1,3-dithiane (**17**) by reaction of **23** with diethyl butyn-3-ylphosphonate [synthesized by coupling 3-(trimethylsilyl)propargyl bromide with diethyl methylphosphonate¹⁹] followed by selective de-ethylation with bromotrimethylsilane.²⁰ **18** and **19** were prepared by treating the corresponding carboxylic acids (**10** and **12**) with excess oxalyl chloride followed by excess diazomethane.²¹

no	R	NMR, δ ppm ²²		mp °C	IC ₅₀ nM
		¹³ C	¹ H		
<u>1</u>	H	--	3.10	148-149	21
<u>2</u>	CH ₃	4.4	2.03	161-162	35
<u>3</u>	CH ₂ CH ₃	13.1, 13.9	1.22, 2.40	130-131	690
<u>4</u>	CH ₂ CH ₂ CH ₃	13.5, 21.4, 22.2	1.03, 1.58, 2.37	133-134	6700
<u>5</u>	CH ₂ OH	51.5	4.48	148-149	76
<u>6</u>	CH ₂ CH ₂ OH	23.8, 61.1	2.67, 3.79	146-147	400
<u>7</u>	CH ₂ CH ₂ CH ₂ OH	15.9, 31.2, 61.5	1.82, 2.51, 3.78	147-148	410
<u>8</u>	CH(OH)CH ₃	24.3, 58.8	1.54, 4.74	153-154	3100
<u>9</u>	C(CH ₃) ₂ OH	31.4, 65.5	1.60	177-178	> 10000
<u>10</u>	CO ₂ H	154.1	--	160 (dec)	7
<u>11</u>	CH ₂ CO ₂ H	26.5, 173.6	3.55	156-161	23
<u>12</u>	CH ₂ CH ₂ CO ₂ H	15.2, 33.2, 177.5	2.69, 2.73	221	5
<u>13</u>	CH ₂ CH ₂ PO ₃ H ₂	14.4, 28.0	2.01, 2.68	185-186	10
<u>14</u>	CO ₂ CH ₃	52.8, 154.3	3.83	181-182	9
<u>15</u>	CH ₂ CO ₂ CH ₂ CH ₃	14.0, 26.6, 61.4, 167.8	1.30, 3.49, 4.22	119-126	460
<u>16</u>	CH ₂ CH ₂ CO ₂ CH ₃	15.4, 33.5, 50.9, 172.3	2.62, 2.72, 3.71	133-135	17
<u>17</u>	CH ₂ CH ₂ P(O)(OCH ₂ CH ₃) ₂	13.4, 16.4, 25.3, 61.6	1.33, 2.07, 2.70, 4.12	102	3000
<u>18</u>	C(O)CHN ₂	60.1, 189.5	5.59	147 (dec)	690
<u>19</u>	CH ₂ CH ₂ C(O)CHN ₂	15.2, 39.5, 54.8, 192.6	2.61, 2.73, 5.35	134 (dec)	830

The new dithianes 3-19 were compared with known compounds 1 and 2 for potency as inhibitors (IC₅₀ = concentration for 50% inhibition) of [³H]EBOB binding using bovine brain membranes.⁶ Addition to 1 of terminal alkane substituents results in progressively lower receptor potency with increasing chain length (2 - 4). In the *n*-alkanol series (5-7) potency drops off with chain length, while secondary and tertiary alcohols (8 and 9) have very low activity. The results with acids 10-13 are very surprising since there are no reports of GABA_A receptor antagonists with acidic functionalities acting at the NCB site. The exceptional potencies of carboxylic acids 10 and 12²³ and phosphonic acid 13 are therefore quite remarkable. Interestingly, there are clusters of arginines and lysines at the channel mouth which provide positively-charged residues that are presumed to serve as anion-concentrating regions normally important for chloride ion flow¹. These findings and earlier structure-activity studies^{24,25} suggest that the *tert*-butyl moiety may enter the channel and approach the NCB site first with the acidic functionality possibly directed towards the mouth of the channel. Esterification of the carboxylic acid moiety reduces the receptor potency 1.3- to 20-

fold (10-12 vs. 14-16) whereas with the phosphonic acid (13) the diethyl phosphonate (17) is 300-fold less active. The high activity of the carboxylic esters implies that suitable linkage to agarose gels or proteins might yield affinity columns or hapten-protein conjugates as immunogens for antibody production. Diazoketones 18 and 19 combine moderate receptor potency with outstanding photoreactivity in pH 7.4 phosphate buffer on irradiation at 254 nm and are therefore candidate photoaffinity probes.

In conclusion, the dithianes with suitably-positioned acidic functional groups retain high receptor potency and form the basis for further derivatizations to obtain photoaffinity probes and affinity columns.

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